



Enhancing Vigor and Viability of Deteriorated True Shallot Seed by Matriconditioning Using Biofertilizer and Washed Rice Water

Muhamad Wahyu Saputra^a, Vega Kartika Sari^a, Slameto^a, Tri Candra Setiawati^b, Ali Wafa^c,
Toni Firmansyah^d, Sundahri^{a,*}

^a Department of Agronomy, Faculty of Agriculture, University of Jember, Jember, Indonesia

^b Department of Soil Science, Faculty of Agriculture, University of Jember, Jember, Indonesia

^c Department of Plant Protection, Faculty of Agriculture, University of Jember, Jember, Indonesia

^d PT East West Seed Indonesia, Jember, Indonesia

Abstract. True Shallot Seed (TSS) is a healthy, cost-effective, and high-yielding alternative planting material instead of shallot bulbs. However, TSS has a short shelf life due to deterioration during storage. The viability and vigor of deteriorated seeds can be enhanced by Plant Growth Promoting Rhizobacteria (PGPR). Washed rice water containing macro- and micronutrients also supports metabolic processes and improves the accumulation of seed nutrients during germination. Therefore, this study aimed to investigate the role of matriconditioning with a mixture of PGPR and washed rice water for enhancing shallot seed vigor and viability. In addition, the physiological quality of shallot seeds in the study had decreased their moisture and germination by 9.09% and 52.25%, respectively. The treatment of Matriconditioning consisted of five levels: Control (M0), Matriconditioning without PGPR (M1), Matriconditioning plus PGPR RhizomaX (M2), Matriconditioning plus PGPR BenprimA (M3), and Matriconditioning plus PGPR FloraOne (M4). Washed rice water consisted of three levels: control (A0), 50% concentration (A1), and 100% concentration (A2). Observed variables were germination rate, maximum growth potential, relative growth rate, uniformity of growth, and vigor index. Data were analyzed by ANOVA, and the Duncan Multiple Range Test was applied for significant results at the 5% level. The results showed that matriconditioning using charcoal and PGPR RhizomaX (M2) could enhance the vigor and viability of deteriorated shallot seeds. The application of washed rice water for five days during germination, especially at a concentration of 100% (A2), also significantly improved the vigor and viability of deteriorated shallot seeds.

Keywords: *Allium cepa* L., germination, seed degeneration, seed priming.

Type of the Paper: Regular Article.

1. Introduction

Shallot is widely used in daily Indonesian consumption, both for health and culinary purposes. According to Shahrajabian et al. [1], shallot is rich in bioactive compounds, such as flavonoids, organosulfur, and phenolic acids. The compounds contained in shallot have antioxidant and anti-inflammatory properties [2]. These compounds also have the potential to inhibit the proliferation of tumor cells, induce apoptosis, and suppress angiogenesis [3]. Furthermore, shallot can reduce the risk of chronic diseases, such as cardiovascular, diabetes, and specific types of cancer [4]. A previous study showed that shallot had the potential as an antimicrobial agent and immunomodulator [5].

The selection of planting material is an important stage for successful shallot cultivation. Shallot bulbs are the most widely used source of cultivation seed among farmers in Indonesia but are prone to seed-borne diseases [6]. A previous study attributed high seed pathogenicity to the unregulated sale of shallot bulbs by farmers and growers, leading to high mutational variations [7]. Therefore, ensuring high-quality seed as planting material is important to prevent additional challenges in shallot cultivation [8].

True Shallot Seed (TSS) has the potential to replace shallot bulbs as a high-quality seed source [9]. Saputri et al. [6] reported significantly lower disease incidence in TSS compared to shallot bulbs. TSS also has lower mutational variations, causing more uniform growth and yield of shallot plants [7]. Atman [10] also reported that the use of TSS reduced seed procurement costs by up to 66.7% and increased harvest yields to 30-34 tons/ha. Therefore, TSS provides an alternative for obtaining quality planting materials in shallot cultivation.

The use of TSS is still uncommon among farmers due to the need for seedling establishment through sowing. A previous study reported that seedling establishment in TSS fails often with low germination rates [7] caused by deterioration during storage. TSS has a relatively short shelf life in tropical regions, less than a year [11]. Rahayu et al. [12] reported the Pancasona variety can be stored for up to 6 months at room temperature. Meanwhile, Yulyatin and Haryati [11] reported that TSS of the Bima variety can only be stored at room temperature for a maximum of 3 months, and an extension beyond this duration will lead to decreasing quality.

Advancing technology is important to provide TSS that maintains high-quality standards after storage. Several strategies can be implemented to achieve high-quality TSS seed, including optimizing production processes [13,14], storage under optimal conditions [12], enhancing quality through pre-planting treatments [15], and adjusting nursery environments [16]. Invigoration is a pre-planting treatment which is a particularly accessible method for farmers to improve seed quality after storage [17].

Matricconditioning (seed priming) is an effective invigoration method for various seed types to improve deterioration [18]. This method can be combined with bioagents, growth regulators, fertilizers, and pesticides [19]. Plant Growth Promoting Rhizobacteria (PGPR) is a commonly used bioagent in seed priming [20]. The use of PGPR in seed priming is reported to enhance viability and vigor of deteriorated seed [20–22]. Additionally, nutrient supplementation can further improve the quality of deteriorated seed by supporting metabolic processes and enhancing nutrient accumulation during germination [23].

Washed rice water is an organic material rich in starch, protein, and various macronutrients and micronutrients [24]. Utami and Hariyanto [25] reported that organic content, such as carbohydrates, minerals, vitamins, and amino acids enhanced seed germination and stimulated

development. Abdelkader et al. [26] also reported that the exogenous addition of amino acids (glutamine, proline, and tryptophan) improved the germination ability of shallot seed. The use of washed rice water as a nutritional source to enhance seed vigor and viability remains uncommon. Therefore, this study aimed to investigate the role of matriconditioning with a mixture of PGPR and rice washing water in enhancing shallot seed vigor and viability.

2. Materials and methods

2.1. Study Site and Materials

This study had been conducted from January 16 to April 8, 2023, in a greenhouse and Seed Technology Laboratory, Faculty of Agriculture, University of Jember. The material used was shallot seed of Lokananta variety, which had substandard quality according to the Minister of Agriculture standards for sowing seed class [27]. Shallot seed has a moisture content of 9.09% and a germination rate of 52.25%. Other materials used include PGPR-biofertilizer RhizomaX (contains PGPR *Rhizobium* sp. 10^{10} cfu/ml, *Bacillus polymixa* 10^{10} cfu/ml, and *Pseudomonas fluorescens* 10^{10} cfu/ml), PGPR-biofertilizer BenprimA (contains PGPR *Bacillus polymixa* 10^7 cfu/ml, dan *Pseudomonas fluorescens* 10^7 cfu/ml), and PGPR-biofertilizer FloraOne (contains PGPR *Rhizobium* sp. $3,4 \times 10^{10}$ cfu/ml, *Pseudomonas fluorescens* $9,3 \times 10^{10}$ cfu/ml, *Azospirillum* sp. $7,3 \times 10^8$, *Aspergillus niger* $3,4 \times 10^7$, and *Tricoderma harzianum* $1,3 \times 10^7$).

2.2. Experimental Design

A split-plot design was adopted with a factorial complete randomized design (CRD) with two factors. The first was matriconditioning treatment with a mixture of PGPR biofertilizer, consisting of 5 levels, namely Control (M0), Matriconditioning using rice husk charcoal (M1), Matriconditioning using rice husk charcoal plus PGPR biofertilizer RhizomaX (M2), Matriconditioning using rice husk charcoal plus PGPR biofertilizer BenprimA (M3), and Matriconditioning using rice husk charcoal plus PGPR biofertilizer FloraOne (M4). The second factor was the application of washed rice water, with 3 levels, namely Control (0% concentration) (A0), 50% concentration (A1), and 100% concentration (A2). Each experiment was repeated 4 times, with each replication containing 100 shallot seeds.

2.3. Seed Treatment

The concentrations of PGPR biofertilizer solutions were 10 g/liter (BenprimA), 10 g/liter (RhizomaX), and 15 ml/liter (FloraOne). A total of 500 g rice was washed with 500 ml well water 4 times, resulting in a stock solution with a concentration of 100%. The stock solution was diluted with well water to make a 50% concentration of washed rice water.

Matriconditioning media were prepared with a mixture of shallot seed, rice husk charcoal, distilled water (control), or PGPR biofertilizer (according to the study treatments). This procedure

also included 3 g, 50 g, 20 ml, and 20 ml of washed rice water, which were applied during germination from day 1 - 5, after planting, with a total of 40 ml used.

2.4. Seed Testing and Observational Variables

The evaluation of vigor and viability of shallot seed had been conducted using the sand test method and incubated in a greenhouse with a daily temperature range of 32-40°C. The testing was carried out on 400 seed samples with 4 replications [28]. The observed variables for vigor and viability were based on previous studies [27,29]. These variables included a) germination rate (%), representing the percentage of normal seedlings after 12 days of planting, b) maximum growth potential (%), including both normal and abnormal seedlings after 12 days of planting, c) relative growth rate (%), showing the percentage of normal seedlings in 24 hours during the 12-day period, d) uniformity of growth (%), representing the percentage of normal seedlings after 7 days of planting, and e) vigor index (%), showing the percentage of normal seedlings at the first count, 6 days after planting.

2.5. Data Analysis

Data analysis was carried out using Analysis of Variance (ANOVA), followed by Duncan's Multiple Range Test (Duncan's test) for significant results at the 5% level. Data analysis was conducted using IBM SPSS Statistics 25.0 software.

3. Results and Discussion

3.1. ANOVA Result

The interaction between matricconditioning and washed rice water application significantly affected the relative growth rate. Matricconditioning significantly affected germination, growth rate, vigor index, uniformity, and maximum growth potential. The application of washed rice water significantly affected germination rate, relative growth rate, vigor index, and uniformity of growth, as shown in Table 1.

Table 1. Results of ANOVA on the effects of matricconditioning and washed rice water application on vigor and viability of TSS seed.

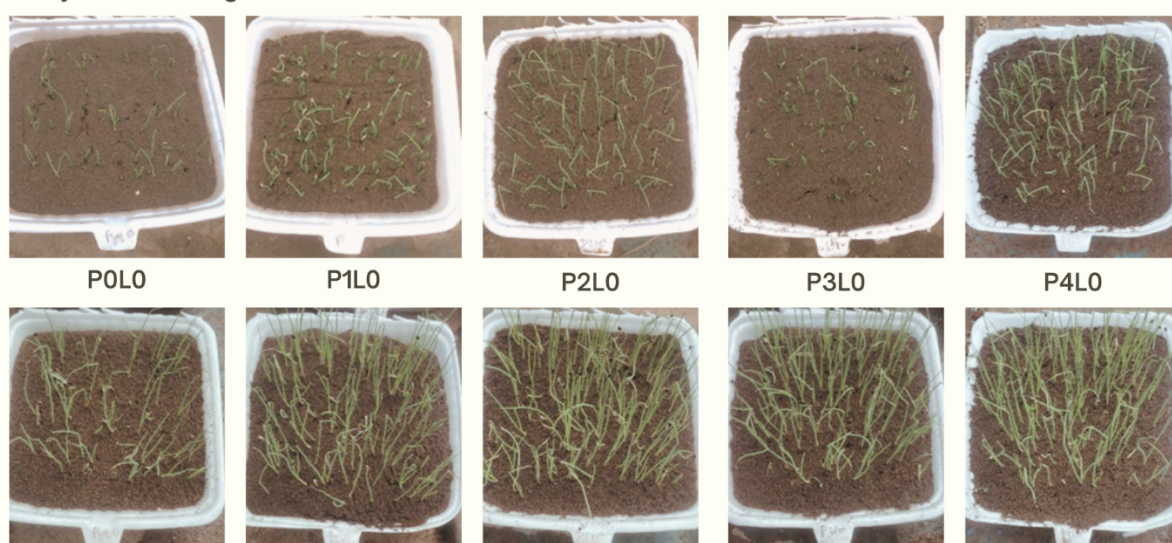
| Observational Variables | F-Value | | | CV |
|--------------------------|----------|----------------------|----------------------|-----|
| | M | A | M*A | |
| Germination rate | 5.0449** | 3.9677* | 2.1165 ^{ns} | 11% |
| Maximum growth potential | 8.8836** | 6.7960** | 2.4535* | 14% |
| Relative growth rate | 6.2932** | 4.5280* | 2.1241 ^{ns} | 16% |
| Uniformity of growth | 6.0337** | 3.4084* | 2.0823 ^{ns} | 12% |
| Vigor index | 4.8535** | 1.5727 ^{ns} | 1.1873 ^{ns} | 11% |
| F 5% | 2.895 | 3.316 | 2.266 | |
| F 1% | 4.500 | 5.390 | 3.173 | |

Notes: * = significant at the 5% level, ** = significant at the 1% level, ns = not significant, M = Matricconditioning, A = Washed rice water; M*A = Interaction of matricconditioning and washed rice water application; CV = Coefficient of variation

3.2. The Effect of Matriconditioning with PGPR on Vigor and Viability of Shallot Seed

The treatment of control (M0) showed the lowest vigor and viability compared to other matriconditioning treatments (Table 2). Seed deterioration causes oxidative stress, leading to damage in proteins, lipids, and deoxyribonucleic acid [30]. This results disrupt hormone activity and seed nutrient mobilization, as reported by a previous study [31]. Seed deterioration was further increased when germination occurred under stressed conditions [32]. Matriconditioning is a priming method that uses a substrate medium to stimulate metabolism and regulate oxygen circulation, optimizing seed germination [19]. Chomontowski et al. [33] reported that solid-matrix priming (matriconditioning) enhanced seed strength and growth rate of *Beta vulgaris* L. Fig. 1 shows the impact of matriconditioning with PGPR on vigor and viability of shallot seed.

6 days after sowing



12 days after sowing

Fig. 1. Shallot seedlings treated with matriconditioning were observed on the 6th and 12th days after sowing.

Table 2. Effect of matriconditioning treatment on vigor and viability of shallot seed

| Matriconditioning | Germination Rate (%) | Relative Growth Rate (%.etmal ⁻¹) | Vigor Index (%) | Uniformity Of Growth (%) | Maximum Growth Potential (%) |
|-------------------|----------------------|---|-----------------|--------------------------|------------------------------|
| M0 | 56.75±10.89 d | 9.61±2.26 c | 38.33±3.19 c | 54.00±3.48 c | 68.58±2.06 c |
| M1 | 67.92±6.90 bc | 12.40±2.00 b | 46.42±3.72 b | 65.33±1.96 bc | 75.42±2.76 bc |
| M2 | 78.17±8.70 a | 15.66±2.31 a | 61.33±5.21 a | 76.08±3.83 a | 88.42±4.09 a |
| M3 | 64.58±11.62 c | 12.12±2.89 b | 45.42±8.89 b | 63.67±8.83 b | 73.33±6.67 bc |
| M4 | 72.25±9.09 ab | 15.26±2.37 a | 57.42±1.65 a | 72.00±4.25 ab | 80.67±2.24 b |

Notes: Data is presented as mean ± standard deviation (n = 12). Values followed by the same letter show no significant difference based on Duncan's analysis at a significance level of 5%

Matriconditioning treatment with a mixture of PGPR biofertilizer Rhizomax (M2) showed the best vigor and viability results compared to other matriconditioning treatments. Hydro-priming can enhance seed respiration, but the effect is relatively low [34]. Seed priming with exogenous phytohormones can further improve seed vigor and viability [35]. Furthermore, PGPR activity includes the production of phytohormones, such as auxins, gibberellins, cytokinins, and abscisic

acid, which enhance seed vigor and viability [20]. PGPR biofertilizer of Rhizomax contains a more complex PGPR content compared to FloraOne. According to a previous study, PGPR bacteria function optimally when operating as a consortium for phytohormone production [36]. However, FloraOne biofertilizer has a more comprehensive PGPR content compared to Rhizomax biofertilizer, resulting in lower vigor and viability of shallot seed. This result was attributed to the presence of *Trichoderma hazarnum* fungi in FloraOne biofertilizer. The use of *Trichoderma* as a seed priming material has been reported to decrease seed germination ability in several horticultural commodities [37].

3.3. The effect of washed rice water treatment on vigor and viability of shallot seed

Treatment A2 resulted in better vigor and viability compared to other washed rice water treatments. The lowest vigor and viability of shallot seed were shown by treatment A1 (50% washed rice water) but were not significantly different from the control (Table 3). Furthermore, the impact of washed rice water on vigor and viability of shallot seed was visualization in Fig. 2. A previous study reported that deteriorated seed often experience a loss of stored nutrients, such as starch, protein, and lipids [38]. Washed rice water contains various organic substances, such as starch, protein, lipids, amylose, and vitamin E [39], as well as various macro and micronutrients [24]. The presence of macro and micronutrients in washed rice water has the potential to enhance the germination ability of deteriorated seed. The priming of seed in inorganic salt solutions, such as NaCl, KCl, KNO₃, K₃PO₄, KH₂PO₄, MgSO₄, and CaCl₂ has been reported to induce seed metabolism during germination. Similarly, the priming of seed with macro and micronutrient ion solutions allows for the adjustment to osmotic pressure, enhances enzyme activity, promotes embryo enlargement, and increases metabolism [23].

Table 3. Effect of washed rice water treatment on vigor and viability of shallot seed

| Washed Rice Water | Germination Rate (%) | Relative Growth Rate (%.etmal ⁻¹) | Vigor Index (%) | Uniformity Of Growth (%) | Maximum Growth Potential (%) |
|-------------------|----------------------|---|-----------------|--------------------------|------------------------------|
| A0 | 66.2±14.08 b | 12.9±3.64 b | 48.8±11.17 b | 64.4±14.02 b | 76.7±9.29 a |
| A1 | 65.75±9.71 b | 12.0±2.73 b | 46.7±8.08 b | 64.2±10.24 b | 75.3±5.72 a |
| A2 | 71.85±10.75 a | 14.1±2.98 a | 53.9±8.59 a | 70.1±11.37 a | 80.0±7.37 a |

Notes: Data is presented as mean ± standard deviation (n = 10). Values followed by the same letter show no significant difference based on Duncan's analysis at a significance level of 5%

The application of washed rice water at a concentration of 100% could significantly improve vigor and viability of shallot seed. However, a concentration of 50% was less effective in enhancing vigor and viability of shallot seed. The mechanism behind the improvement was more clearly influenced by the presence of macro and micronutrients. Although washed rice water was reported to contain a lot of starch [39], no study has reported that adding exogenous starch can enhance seed germination ability. Furthermore, exogenous glucose in seed, when present in sufficiently high concentrations, can induce abscisic acid hormone activity, leading to dormancy

[40,41]. The addition of exogenous glucose at low concentrations (0.1 – 0.5 mM) can increase seed germination ability under saline stress conditions [42].

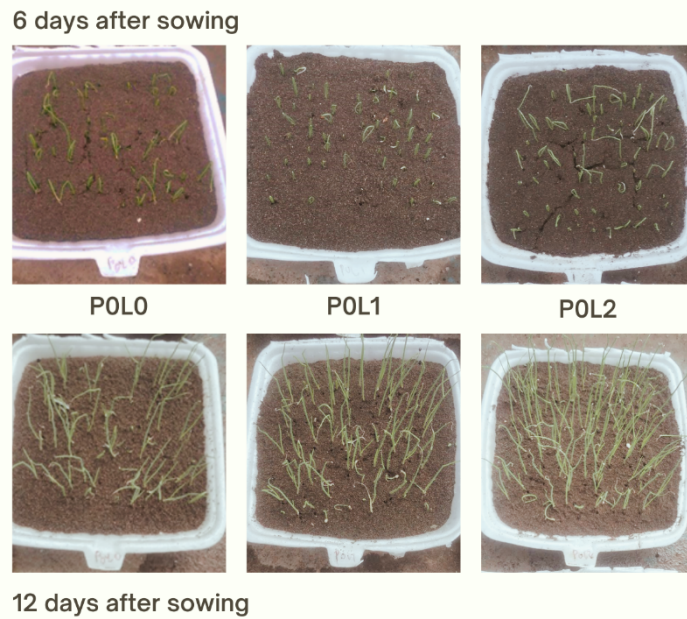


Fig. 2. Shallot seedlings treated with washed rice water were observed on the 6th and 12th days after sowing.

Table 4. Effect of the interaction between matriconditioning with PGPR and washed rice water treatment on vigor and viability of shallot seed

| Matriconditioning | Washed Rice Water | Germination Rate (%) | Relative Growth Rate (%.etmal ⁻¹) | Vigor Index (%) | Uniformity Of Growth (%) | Maximum Growth Potential (%) |
|-------------------|-------------------|----------------------|---|-----------------|--------------------------|------------------------------|
| M0 | A0 | 55.25±12.79a | 9.52±1.94gh | 37.75±12.97a | 52.75±13.16a | 67.25±12.17a |
| | A1 | 53.25±6.180a | 8.46±0.95h | 34.75±7.41a | 50.50±7.23a | 67.00±6.75a |
| | A2 | 61.75±10.69a | 10.85±2.78fg | 42.5±13.96a | 58.75±13.05a | 71.5±10.36a |
| M1 | A0 | 68.25±7.50a | 12.77±1.78cdef | 50.50±9.40a | 65.75±7.08a | 78.75±10.85a |
| | A1 | 65.00±4.53a | 11.48±1.67cfe | 41.50±9.75a | 62.75±3.34a | 72.00±5.79a |
| | A2 | 70.50±7.12a | 12.97±2.17cdef | 47.25±9.22a | 67.5±6.34a | 75.50±8.41a |
| M2 | A0 | 76.00±6.75a | 15.66±1.77ab | 61.75±4.43a | 73.25±5.63a | 88.75±4.97a |
| | A1 | 75.50±10.23a | 14.06±2.25bcd | 54.75±9.64a | 73.50±10.62a | 83.25±10.23a |
| | A2 | 83.00±6.44a | 17.25±1.66a | 67.50±7.33a | 81.50±5.02a | 93.25±3.77a |
| M3 | A0 | 53.50±9.63a | 9.53±2.070gh | 34.25±10.72a | 52.25±8.14a | 64.75±8.64a |
| | A1 | 65.75±3.63a | 12.02±2.11def | 46.00±12.94a | 65±3.81.00a | 74.25±6.26a |
| | A2 | 74.50±8.73a | 14.81±1.55abc | 56.00±8.98a | 73.75±8.07a | 81.00±10.98a |
| M4 | A0 | 78.00±10.63a | 17.03±2.06a | 59.75±8.73a | 78.00±10.63a | 83.75±9.55a |
| | A1 | 69.25±5.54a | 13.93±1.84bcde | 56.25±8.02a | 69.25±5.54a | 79.75±6.65a |
| | A2 | 69.50±7.40a | 14.8±2.030abc | 56.25±9.91a | 68.75±8.04a | 78.50±7.26a |

Notes: Data is presented as mean ± standard deviation (n = 4). Values followed by the same letter show no significant difference based on Duncan's analysis at a significance level of 5%

3.4. Effect of the interaction between matriconditioning with PGPR and washed rice water treatment on vigor and viability of shallot seed

The control treatment M0A0 showed the lowest relative growth rate compared to others (Table 4). The interaction between matriconditioning and washed rice water treatment caused varied vigor and viability of shallot seed. Treatment M3A0 showed the lowest results in germination rate, vigor index, uniformity, and maximum growth potential, but was not significantly different from the control treatment (M0A0). However, interactions between M3A1

and M3A2 produced better vigor and viability of shallot seed compared to M0A1 and M0A2.

The interaction between matriconditioning with a mixture of PGPR biofertilizer and washed rice water showed different responses. M3A2 exhibited the best vigor and viability compared to M3A0 and M3A1. However, M4A0 showed the best results in vigor and viability of shallot seed compared to M4A1 and M4A2. The combination of washed rice water and PGPR biofertilizer enables the formation of a complex bacterial consortium that could influence the activity of rhizobacteria as plant growth-promoting agents through mechanisms, such as nitrogen fixation, phytohormone production, phosphorus, and potassium solubilization [43]. Non-fermented washed rice water contains several strains of rhizobacteria from the groups *Pantoea*, *Klebsiella*, *Bacillus*, *Enterobacter*, and *Stenotrophomonas* [24]. Furthermore, the complexity of PGPR bacterial population in matriconditioning treatment with a mixture of PGPR biofertilizer and washed rice water may lead to competition among rhizobacteria for nutrients, space, and environmental factors. According to Dhungana and Itoh [44], the majority of rhizobacteria have the ability to produce IAA (indole-3-acetic acid), while others play a role in degrading IAA in plants.

4. Conclusions

In conclusion, both matriconditioning and washed rice water treatments, particularly when applied individually, improved the vigor and viability of deteriorated shallot seeds. The best outcome was observed with matriconditioning using rice husk media combined with the PGPR biofertilizer Rhizomax. Additionally, treating the seeds with 100% concentration of washed rice water for 5 days also significantly enhanced shallot seed vigor and viability.

Data Availability Statement

Data will be shared upon request by the readers.

CRedit Authorship Contribution Statement

M.W.S: Conceptualization, Data curation, Investigation, Methodology, Project administration, Writing – original draft. V.K.S.: Validation, Writing – review & editing, Project administration. I.F.: Methodology, Validation, Writing – review & editing, administration. SL: Methodology, Project administration, Resources, Validation, Visualization, Writing – review & editing. T.C.C: Validation, Writing – review & editing, Validation. A.W.: Validation, Writing – review & editing, Visualization. S.H.: Supervision, Funding acquisition, Validation, review & editing, Project administration, Software, Validation, Visualization, Writing original draft.

Declaration of Competing Interest

The authors declare no known competing financial interests or personal relationships that could have influenced this study.

Acknowledgement

The author(s) declare that financial support was received for the research, authorship, and/or publication of this article. The authors gratefully acknowledged research supporting research program of Institute for Research and Community Service, University of Jember for funding this study and PT EAST WEST SEED INDONESIA (EWINDO) for collaborating with us so that this research could be completed.

References

- [1] Shahrajabian MH, Sun W, Cheng Q. Chinese onion, and shallot, originated in Asia, medicinal plants for healthy daily recipes. *Not Sci Biol* 2020;12:197–207. <https://doi.org/10.15835/nsb12210725>.
- [2] Ounjaijean S, Chachiyo S, Kulprachakarn K, Boonyapranai K, Srichairatanakool S, Rerkasem K. Antioxidant and Anti-inflammatory Protective Properties of Thai Shallot (*Allium ascalonicum* cv. Chiangmai) Juice on Human Vascular Endothelial Cell Lines (EA.hy926). *Walailak Journal of Science and Technology (WJST)* 2019;16:175–84. <https://doi.org/10.48048/wjst.2019.6222>.
- [3] Forma A, Chilimoniuk Z, Januszewski J, Sitarz R. The Potential Application of Allium Extracts in the Treatment of Gastrointestinal Cancers. *Gastroenterol Insights* 2021;12:136–46. <https://doi.org/10.3390/gastroent12020012>.
- [4] Chakraborty AJ, Uddin TM, Matin Zidan BMR, Mitra S, Das R, Nainu F, et al. Allium cepa: A Treasure of Bioactive Phytochemicals with Prospective Health Benefits. *Evidence-Based Complementary and Alternative Medicine* 2022;2022:1–27. <https://doi.org/10.1155/2022/4586318>.
- [5] Zhao X-X, Lin F-J, Li H, Li H-B, Wu D-T, Geng F, et al. Recent Advances in Bioactive Compounds, Health Functions, and Safety Concerns of Onion (*Allium cepa* L.). *Front Nutr* 2021;8. <https://doi.org/10.3389/fnut.2021.669805>.
- [6] Saputri AS, Tondok ET, Hidayat SH. Insidensi Virus dan Cendawan pada Biji dan Umbi Bawang Merah. *Jurnal Fitopatologi Indonesia* 2019;14:222. <https://doi.org/10.14692/jfi.14.6.222>.
- [7] Rosliani R, Waluyo N, Yufdy MP, Harmanto, Sulastrini I, Handayani T, et al. Benih Biji Bawang Merah (True Seed of Shallot) di Indonesia. Jakarta: IAARD Press; 2022. <http://repository.pertanian.go.id/handle/123456789/16142>
- [8] Adiyoga W. Seed Systems in the Four Shallot Producing Areas of Java: A Focus Group Discussion. *E3S Web of Conferences* 2021;232:01003. <https://doi.org/10.1051/e3sconf/202123201003>.
- [9] Rabinowitch HD. Shallot (*Allium cepa* L. *Aggregatum* Group) Breeding. In: Al-Khayri JM, Jain SM, Johnson D V, editors. *Advances in Plant Breeding Strategies: Vegetable Crops*, Cham: Springer International Publishing; 2021, p. 99–154. https://doi.org/10.1007/978-3-030-66965-2_3.
- [10] Atman A. Teknologi Budidaya Bawang Merah Asal Biji. *Jurnal Sains Agro* 2021;6:11–21. <https://doi.org/10.36355/jsa.v6i1.497>.
- [11] Yulyatin A, Haryati Y. Pengujian Daya Berkecambah Biji Bawang Merah Selama 7 Periode Simpan. *BPTP Jawa Barat* 2016;6:5–8. <https://repository.pertanian.go.id/handle/123456789/6573>
- [12] Rahayu A, Waluyo N, Azmi C. Pengaruh Lama dan Ruang Simpan terhadap Perkecambahan Benih True Shallot Seed (TSS). *Peningkatan Produktivitas Pertanian Era Society 5.0 Pasca Pandemi*, Politeknik Negeri Jember; 2021, p. 244–54. <https://doi.org/10.25047/agropross.2021.227>.

- [13] Fahrianty D, Poerwanto R, Widodo WD, Palupi ER. Improvement of Flowering and Seed Yield of Shallot Variety Bima through Vernalization and Application of GA3. *Jurnal Ilmu Pertanian Indonesia* 2020;25:245–52. <https://doi.org/10.18343/jipi.25.2.245>.
- [14] Widiarti W, Wijaya I, Umarie I. Optimalisasi Teknologi Produksi True Shallot Seed (Biji Biologi) Bawang Merah (*Allium ascalonicum* L). *Agritrop* 2017;15:203–16. <http://jurnal.unmuhsember.ac.id/index.php/AGRITROP/article/view/1174>
- [15] Sudjarwo HK, Moeljani IR, Pribadi DU. Pengaruh Lama Perendaman GA3 dan Beberapa Macam TSS Terhadap Pertumbuhan Tanaman Bawang Merah (*Allium ascalonicum* L.). *Jurnal Ilmu-Ilmu Pertanian Indonesia* 2021;23:129–35. <https://doi.org/10.31186/jipi.23.2.129-135>.
- [16] Azmi C, Rahayu A, Rosliani R, Hermanto C. Pertumbuhan Benih True Shallot Seed (TSS) pada Berbagai Media Semai. In: Matra DD, Devy NF, editors. *Prosiding Seminar Nasional PERHORTI 2020 "Sinergisme Membangun Kawasan Hortikultura Tangguh dan Menyehatkan"*, Malang: Perhimpunan Hortikultura Indonesia (PERHORTI); 2020, p. 234–45. https://www.researchgate.net/publication/353174901_Pertumbuhan_Benih_True_Shallot_Seed_TSS_pada_Berbagai_Media_Semai
- [17] Ranganathan U, Groot SPC. Seed Longevity and Deterioration. In: Dadlani M, Yadava DK, editors. *Seed Science and Technology*, Singapore: Springer Nature Singapore; 2023. <https://doi.org/10.1007/978-981-19-5888-5>.
- [18] Panda D, Mondal S. Seed Enhancement for Sustainable Agriculture: An Overview of Recent Trends. *Plant Arch* 2020;20:2320–32. <http://www.plantarchives.org/List%20SI%2020,%20SUPP-1,2020.html>
- [19] Pagano A, Macovei A, Xia X, Padula G, Holubowicz R, Balestrazzi A. Seed Priming Applied to Onion-Like Crops: State of the Art and Open Questions. *Agronomy* 2023;13:288. <https://doi.org/10.3390/agronomy13020288>.
- [20] Kenneth OC, Nwadike EC, Kalu AU, Unah UV. Plant Growth Promoting Rhizobacteria (PGPR): A Novel Agent for Sustainable Food Production. *Am J Agric Biol Sci* 2019;14:35–54. <https://doi.org/10.3844/ajabssp.2019.35.54>.
- [21] Ha-Tran DM, Nguyen TTM, Hung S-H, Huang E, Huang C-C. Roles of Plant Growth-Promoting Rhizobacteria (PGPR) in Stimulating Salinity Stress Defense in Plants: A Review. *Int J Mol Sci* 2021;22:3154. <https://doi.org/10.3390/ijms22063154>.
- [22] Sundahri S, Mursyidto T, Setiawati TC, Susilo HA, Wafa A. Inducing The Viability of Deteriorated Jatropha Seed Through Matriconditioning Technology and Pseudomonas Fluorescens as Biological Agent. *Devotion : Journal of Research and Community Service* 2023;4:1352–73. <https://doi.org/10.59188/devotion.v4i6.502>.
- [23] Martínez-Ballesta M del C, Egea-Gilabert C, Conesa E, Ochoa J, Vicente MJ, Franco JA, et al. The Importance of Ion Homeostasis and Nutrient Status in Seed Development and Germination. *Agronomy* 2020;10:504. <https://doi.org/10.3390/agronomy10040504>.
- [24] Nabayi A, Sung CTB, Zuan ATK, Paing TN, Akhir NIM. Chemical and Microbial Characterization of Washed Rice Water Waste to Assess Its Potential as Plant Fertilizer and for Increasing Soil Health. *Agronomy* 2021;11:2391. <https://doi.org/10.3390/agronomy11122391>.
- [25] Utami ESW, Hariyanto S. Organic Compounds: Contents and Their Role in Improving Seed Germination and Protocorm Development in Orchids. *International Journal of Agronomy* 2020;2020:1–12. <https://doi.org/10.1155/2020/2795108>.
- [26] Abdelkader M, Voronina L, Puchkov M, Shcherbakova N, Pakina E, Zargar M, et al. Seed Priming with Exogenous Amino Acids Improves Germination Rates and Enhances Photosynthetic Pigments of Onion Seedlings (*Allium cepa* L.). *Horticulturae* 2023;9:80. <https://doi.org/10.3390/horticulturae9010080>.
- [27] Menteri Pertanian Republik Indonesia. *Pedoman Teknik Sertifikasi Benih Bawang Merah*. Indonesia; 2017.

- https://fungsional.pertanian.go.id/ujikompij/assets/file/elearning/elearning_72_5f22855bd80e3.pdf
- [28] Direktorat Perbenihan Hortikultura. Pedoman Uji Mutu Laboratorium. Jakarta: Direktorat Jenderal Hortikultura Kementerian Pertanian; 2016. https://fungsional.pertanian.go.id/ujikompij/assets/file/elearning/elearning_72_5f22872c06bb4.pdf
- [29] Ingmar O, Setiyono S, Savitri DA, Novijanto N. Effect of Seed Coating and Packaging Material on Viability and Vigor of Soybean Seed in Room Temperature Storage. *Journal of Applied Agricultural Science and Technology* 2023;7:109–18. <https://doi.org/10.55043/jaast.v7i2.127>.
- [30] Adetunji AE, Adetunji TL, Varghese B, Sershen, Pammenter NW. Oxidative Stress, Ageing and Methods of Seed Invigoration: An Overview and Perspectives. *Agronomy* 2021;11:2369. <https://doi.org/10.3390/agronomy11122369>.
- [31] Joo EH, Kim YR, Kim N, Jung JE, Han SH, Cho HY. Effect of Endogenic and Exogenic Oxidative Stress Triggers on Adverse Pregnancy Outcomes: Preeclampsia, Fetal Growth Restriction, Gestational Diabetes Mellitus and Preterm Birth. *Int J Mol Sci* 2021;22:10122. <https://doi.org/10.3390/ijms221810122>.
- [32] Wang Y, Jiang W, Cheng J, Guo W, Li Y, Li C. Physiological and Proteomic Analysis of Seed Germination under Salt Stress in Mulberry. *Frontiers in Bioscience-Landmark* 2023;28:49. <https://doi.org/10.31083/j.fbl2803049>.
- [33] Chomontowski C, Wzorek H, Podlaski S. Impact of sugar beet seed priming on seed quality and performance under diversified environmental conditions of germination, emergence and growth. *J Plant Growth Regul* 2020;39:183–9. <https://doi.org/10.1007/s00344-019-09973-2>.
- [34] El-Mergawi RA, Abd El-Wahed MSA. Effect of exogenous salicylic acid or indole acetic acid on their endogenous levels, germination, and growth in maize. *Bull Natl Res Cent* 2020;44:167. <https://doi.org/10.1186/s42269-020-00416-7>.
- [35] Kosakivska I V, Vedenicheva NP, Babenko LM, Voytenko L V, Romanenko KO, Vasyuk VA. Exogenous phytohormones in the regulation of growth and development of cereals under abiotic stresses. *Mol Biol Rep* 2022;49:617–28. <https://doi.org/10.1007/s11033-021-06802-2>.
- [36] Mudi L, Sutariati GAK, Hamriani, Roby. Aplikasi Konsorsium Endo-Rhizobakteri Untuk Meningkatkan Vigor Benih Padi Gogo Lokal Consortium. *Jurnal Agrotech* 2021;11:1–7. <https://doi.org/10.31970/agrotech.v11i1.61>.
- [37] Sánchez-Montesinos B, Diánez F, Moreno-Gavira A, Gea FJ, Santos M. Role of *Trichoderma aggressivum* f. *europaeum* as Plant-Growth Promoter in Horticulture. *Agronomy* 2020;10:1004. <https://doi.org/10.3390/agronomy10071004>.
- [38] Farooq MA, Ma W, Shen S, Gu A. Underlying Biochemical and Molecular Mechanisms for Seed Germination. *Int J Mol Sci* 2022;23:8502. <https://doi.org/10.3390/ijms23158502>.
- [39] Pereira C, Lourenço VM, Menezes R, Brites C. Rice Compounds with Impact on Diabetes Control. *Foods* 2021;10:1992. <https://doi.org/10.3390/foods10091992>.
- [40] Price J, Li T-C, Kang SG, Na JK, Jang J-C. Mechanisms of Glucose Signaling during Germination of *Arabidopsis*. *Plant Physiol* 2003;132:1424–38. <https://doi.org/10.1104/pp.103.020347>.
- [41] Yuan K, Wysocka-Diller J. Phytohormone signalling pathways interact with sugars during seed germination and seedling development. *J Exp Bot* 2006;57:3359–67. <https://doi.org/10.1093/jxb/erl096>.
- [42] Hu M, Shi Z, Zhang Z, Zhang Y, Li H. Effects of exogenous glucose on seed germination and antioxidant capacity in wheat seedlings under salt stress. *Plant Growth Regul* 2012;68:177–88. <https://doi.org/10.1007/s10725-012-9705-3>.

- [43] Denaya S, Yulianti R, Pambudi A, Effendi Y. Novel microbial consortium formulation as plant growth promoting bacteria (PGPB) agent. IOP Conf Ser Earth Environ Sci 2021;637:012030. <https://doi.org/10.1088/1755-1315/637/1/012030>.
- [44] Dhungana SA, Itoh K. Effects of Co-Inoculation of Indole-3-Acetic Acid-Producing and -Degrading Bacterial Endophytes on Plant Growth. Horticulturae 2019;5:17. <https://doi.org/10.3390/horticulturae5010017>.